THE DEVELOPMENT OF MULTIFUNCTIONAL FLUORESCENT PROBES OF RHODAMINE-PYRIMIDINE TYPE

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Introduction

6(1,1,1)

6(1,2,1)

6(1,1,2)

6(1,2,2)

6(1,1,3)

6/1 2 2)

One of the most important organelle studied with respect to the cell viability is mitochondrion. The fluorescent visualization represents a convenient method for study of processes in the mitochondria.¹ Although several fluorescent systems for mitochondria visualization were developed there are still some gaps that remain unsolved.²³ This report is focused on the development of fluorescent systems suitable for staining the mitochondria, using cheap and highly accessible Rhodamine B. The method employs the solid-phase chemistry with application of combinatorial approach which allows the high throughput synthesis for systematic study of fluorescent properties of prepared derivatives. The studied fluorescent systems are formed by central core, unit for control of permeability or solubility and a fluorescent dye bound via appropriate linker (Figure 1).



(i) Fmoc-Ala-OH, *N*-hydroxybenzotriazole (HOBt), DMAP, DIC, DMF/DCM (1:1), 3 h; (ii) 50% piperidine, DMF, 15 min; (iii) Fmoc-Phe-OH, HOBt, DIC, DMF/DCM (1:1), 3 h; (iv) 4,6-dichloro-5-nitro-pyrimidine, DIEA, dry DMF, 2 h; (v) phenylenediamine, DIEA, dry DMF, 5 h; (vi) Rhodamine B, HOBt, DIC, DMF/DCM (1:1), 16 h; (vii) SnCl₂.2H₂O, DIEA, DMF, N₂, rt, 16 h or Na₂S₂O₄, K₂CO₃, TBAHS, H₂O/DCM, rt, 16 h; (viii) 50% TFA in DCM, rt, 1 h.

				*					
6(2,1,1)	1(2]	NO ₂	р-	83	25	559	580	0.1809	yes
6(2,2,1)	1(2)	NH_2	<i>p</i> -	59	32	350	475	0.0067	yes
6(2,1,2)	1(2)	NO ₂	<i>m</i> -	73	37	558	582	0.3845	yes
6(2,2,2)	1(2)	NH_2	m-	62	23	550	568	0.1238	yes
6(2,1,3)	1(2)	NO ₂	0-	61	5	560	582	0.0373	yes
6(2,2,3)	1(2)	$\rm NH_2$	0-	69	11	336	463	0.0052	yes
11(1,1,1)	1(1)	NO ₂	<i>p</i> -	70	70	555	580	0.2712	yes
11(1,2,1)	1(1)	NH_2	<i>p</i> -	80	23	553	583	0.3019	no
11(1,1,2)	1(1)	NO ₂	<i>m</i> -	89	92	560	586	0.1841	yes
11(1,2,2)	1(1)	NH ₂	<i>m-</i>	56	14	557	582	0.2149	no
11(1,1,3)	1(1)	NO ₂	0-	72	76	568	592	0.0528	yes
11(1,2,3)	1(1)	$\rm NH_2$	0-	77	67	568	590	0.0284	yes
11(2,1,1)	1(2)	NO ₂	<i>p</i> -	69	14	560	590	0.2562	yes
11(2,1,2)	1(2)	NO ₂	<i>m</i> -	78	22	566	590	0.0995	yes
11(2,1,3)	1(2)	NO ₂	0-	68	35	350	489	0.0064	no
11(2,2,1)	1(2)	NH ₂	<i>p</i> -	55	4	568	590	0.2549	yes
11(2,2,2)	1(2)	NH ₂	<i>m</i> -	51	7	373	439	0.0233	yes
11(2,2,3)	1(2)	NH_2	0-	72	15	562	586	0.3125	yes

82

77

82

65

85

67

 NO_2

 NH_2

 NO_2

 $\rm NH_2$

 NO_2

p-

р-

m-

61

36

86

30

83

0.2132

0.6912

0.0011

0.0313

0.2254

no

no

yes

yes

yes

580

572

591

459

581

556

550

566

393

557

[°]Purity of crude products estimated from LC traces. ^bIsolated yield after HPLC purification.

The basic fluorescent properties

All compounds were tested for their basic fluorescent properties in DMSO. The excitation and emission spectra of prepared compounds mostly showed similar shape and maxima as Rhodamine B. The various substitutions of central group as well as different linker affect significantly the fluorescence intensity. Interestingly some derivatives do not exhibited rhodamine excitation maximum (around 560 nm), but only one maximum between 336-393 nm (Table 1). The emission intensity of this compounds at rhodamine excitation was much more higher when the measurement was done in water (Figure 2).

pH stability and photostability

The pH dependence study was performed for derivatives **6(1,1,2)**, **6(2,1,3)**, **(1,1,3)** and **11(2,1,3)**. The fluorescence emission measured under various pH is demonstrated in



Figure 3: Dependence of fluorescence intensity on pH for compounds $6(1,1,2) \blacktriangle, 6(1,1,3) \blacklozenge, 6(2,1,3) \bigcirc$ and $11(2,1,3) \blacksquare$.

The photostability of selected compounds was tested. During one hour there was no decreasing of intensite in all cases (Figure 4).



Figure 4: Intensity of fluorescence of derivatives **6(1,1,2)**, **6(1,1,3)**, **6(2,1,3)** and **11(2,1,3)** during one hour measurement. **6(1,1,2)**, black; **6(1,1,3)**, blue; **6(2,1,3)**, green; **11(2,1,3)**, red.

Penetration the cell membrane

The prepared compounds were studied for the ability to penetrate to the living cells (U-2-OS human osteosarcoma). From the 24 studied compounds 18 of them were able to penetrate into the cell interior (Table 1). According to the fluorescent microscopy observation all of them were selectively accumulated in mitochondria (Figure 5).

b



Figure 2: Fluorescence intensity of derivative **6(1,2,3)** according to water/DMSO ratio. Excitation wavelength is 560 nm.

	$\lambda_{ex}/\lambda_{em}(QY)$					
Compound	DMSO	Water				
6(1,1,2)	393/459 (0,03)	0,0076				
6(1,1,3)	386/463 (0,0871)	0,2997				
6(2,2,2)	350/475 (0,0067)	0,1862				
6(2,1,3)	336/463 (0,0052)	0,1024				
11(2,1,3)	350/489 (0,0064)	0,1928				
11(2,2,3)	373/439 (0,0233)	0,3603				



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Figure 5: Fluorescent image of the compound with ability to penetrate to U-2-OS cell: a) NADH autofluorescence at 355 nm visualizing live mitochondria; b) cells after 15 min. treatment with **11(2,2,3)** excited at 561 nm; c) overlay of (a) and (b)

References:

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